

# Eucalyptus oil nanoemulsion-impregnated chitosan film: antibacterial effects against a clinical pathogen, *Staphylococcus aureus*, in vitro

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**Abstract:** Eucalyptus oil (*Eucalyptus globulus*) nanoemulsion was formulated using low- and high-energy emulsification methods. Development of nanoemulsion was optimized for system parameters such as emulsifier type, emulsifier concentration, and emulsification methods to obtain a lower droplet size with greater stability. The minimized droplet diameter was achieved using the high-energy method of ultrasonication. Tween 80 was more effective in reducing droplet size and emulsion appearance when compared to Tween 20. Stable nanoemulsion was formulated with Tween 80 as a surfactant, and the particle size was found to be 9.4 nm (1:2 v/v). The eucalyptus oil nanoemulsion was impregnated into chitosan (1%) as a biopolymer in varying concentrations. Further, the film was characterized by moisture content, microscopic study, X-ray diffraction, and Fourier transform infrared spectroscopy. Also, the film with and without nanoemulsion was evaluated against *Staphylococcus aureus*. The nanoemulsion-impregnated chitosan film showed higher antibacterial activity than chitosan film. These results support the inclusion of nanoemulsion-impregnated chitosan film in wound management studies.

**Keywords:** essential oil, emulsion, biopolymer, impregnation, thin film, wound isolate

## Introduction

Wounds are physical injuries that occur due to an accident that results in a breaking or opening of the skin. The exposed tissue leads to a moist and natural medium for microbial growth and proliferation. The bacterial strain *Staphylococcus aureus*, a Gram-positive opportunistic bacterium, is found to be present in skin abrasions and burns and can lead to infection and sepsis.<sup>1</sup> The process of wound healing is a complex and dynamic healing process that involves three important phases: inflammation, proliferation, and tissue remodeling.<sup>2,3</sup>

The use of essential oils has been reported for human diseases for centuries because they contain enriched bioactive compounds of medicinal value.<sup>4</sup> In recent years, the majority of the world's population now depend on traditional medicine because of the eco-friendliness and cost-effective production.<sup>5</sup> The permeation of essential oil compounds is faster in the skin due to their volatile nature and fat-soluble substances present in them.<sup>6,7</sup> The plant-based essential oils have been shown to possess antimicrobial, wound-healing, insecticidal, and pharmaceutical properties.<sup>2</sup>

For application studies, essential oil-based emulsion formation is important due to the hydrophobic nature of the oil system, and it is necessary to formulate a soluble nature using an emulsification technique. Emulsions are mixtures of two immiscible liquids that are stabilized by a surfactant. Emulsion droplets that are extremely small in

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size, ranging from 20–200 nm, are called nanoemulsions. To prepare nanoemulsions, the high-energy methods are widely employed; these include microfluidization, high-pressure homogenization, and ultrasonication. The method that is most widely used is ultrasonication, because it is economical and easy to use.<sup>8–10</sup> The antimicrobial oil-in-water nanoemulsion was shown to possess a broad spectrum of activity against bacterial pathogens due to the reduced droplet diameter.<sup>11,12</sup> Recently, by our group, the ultrasonic emulsification of eucalyptus oil nanoemulsion was studied, and size reduction was achieved with the aid of ultrasonication time. Also, the stabilized emulsion system was studied for topical application against *S. aureus* by in vitro and was shown to be nonirritant, and high wound healing activity was observed in Wistar rats in an in vivo experiment.<sup>13</sup>

The new generation of wound dressings is the product of the combination of antibacterial agents onto/into the biodegradable polymer material. Efforts are made to ensure a sustained release of active compound into the wounded site so that the infection is highly reduced. An effective means of remedy, at the wounded site is the use of antimicrobial bandages/scaffolds with which the active compound is used in minimal amounts and ensures that the healing activity is efficient at that site.<sup>14</sup>

In recent years, application studies have focused on biodegradable films made from natural polymers in pharmaceutical and food industries. Such polymers may be proteins of plant or animal origin and biological materials. Their chemical nature determines the enhancement of physical and mechanical properties of the resulting film.<sup>15–17</sup> The use of synthetic chemicals such as polyvinyl alcohol and polyvinylpyrrolidone based films had developed resistance in microbes, and also was not applicable for a longer usage in the environment due to non degradable properties. The use of plastic-based products has become part of our life and led to the rapid emergence of the plastic industry in the past several decades. But the films made of synthetic polymers are non-biodegradable and lead to serious ecological problems, resulting in environmental pollution.<sup>18</sup>

Chitosan is the second most abundant polysaccharide from arthropod exoskeletons. It is a nontoxic, biodegradable, antibacterial, chelating biopolymer produced by the deacetylation of chitin, an abundant organic resource.<sup>19–21</sup> Natural biopolymeric films with essential oil have potential advantage over the synthetic products since they are totally biodegradable and are derived from biological materials.<sup>22</sup> Particularly in biomaterials research, they have been found to have enhanced healing rates and tissue-repair strengths.<sup>23–25</sup> The casting solution of chitosan with bioactive compounds

have been widely studied via the incorporation of drugs, proteins, dyes,<sup>26–28</sup> essential oils,<sup>29,30</sup> and phenolic compounds.<sup>31</sup> Chitosan films have been incorporated with thyme oil for potential wound-healing applications.<sup>32</sup> In most studies, a nonionic surfactant was used in essential oil film-forming solution to achieve solubility of oil and to make homogeneous film solution.<sup>33–35</sup> Instead, these can be formulated into emulsion based film formation to have more stability and homogeneous solution.

The present study was focused on impregnating eucalyptus oil nanoemulsion into chitosan to develop a film. Further, this was studied for physicochemical properties through moisture content, microscopic studies, X-ray diffraction, and Fourier transform infrared (FTIR) spectroscopy studies. Finally, the obtained nanoemulsion-impregnated chitosan film (NE-CH) was evaluated against *S. aureus* by in vitro studies such as agar disc diffusion and plate count method.

## Materials and methods

### Materials

Chitosan powder (90% deacetylated), Tween 80, Tween 20, and acetic acid were obtained from Sigma-Aldrich Co. (St Louis, MO, USA). Eucalyptus oil and nutrient agar were purchased from HiMedia (Mumbai, India). Ultrapure water obtained using a Cascada™ Biowater System with a resistivity not less than 18.2 MΩ·cm<sup>-1</sup> was used for the preparation of all solutions. All other chemicals used were of analytical reagent grade.

### Methods

#### Formulation of eucalyptus oil nanoemulsion

Oil-in-water nanoemulsion was prepared by the procedure previously reported.<sup>36</sup> Briefly, oil-in-water nanoemulsion was formulated using eucalyptus oil (6%), nonionic surfactant (Tween 80; Tween 20), and water (1:2 v/v). Initially, the emulsion was prepared by adding water to oil and surfactant mixture using a magnetic stirrer. Then, the coarse mixture was subjected to ultrasonic emulsification using a 20 kHz sonicator with a maximum power output of 750 W.

#### Preparation of NE-CH

Chitosan solution (1% w/v) was prepared using 1% acetic acid. After complete mixing, the solution was filtered with Whatman number 1 paper and used for further studies. Different percentages of eucalyptus oil nanoemulsion (0%, 1%, 3%, and 5% v/v) and chitosan solution were mixed, and this was homogenized using a sonicator for 10 minutes. The film solutions were cast over the evenly leveled acrylic plates and dried at 50–60°C and 60%±2% relative humidity for 48 hours.

### Phase contrast microscopy

The surfaces of the films with and without nanoemulsion were observed with a phase contrast microscope coupled with a camera (Leica DM-2500). The pictures were taken at different magnifications.

### Moisture content analysis

Films with and without nanoemulsion loaded (3×3 cm) were weighed before and after drying in an oven at 105°C for 24 hours.<sup>15</sup> Moisture content was calculated as follows: moisture content =  $M_0 - M_f / M_f$  where  $M_0$  was the initial film weight (mg) and  $M_f$  was the final weight (mg). Moisture content was expressed as milligrams water per milligrams dry solids.

### X-ray diffraction studies

The X-ray diffractograms of the chitosan and nanoemulsion films were characterized with a powdered X-ray diffractometer (model: D8 Advance; Bruker Optik GmbH, Ettlingen, Germany) using Ni-filtered radiation. About 1×1 cm of the film sample was deposited on the sample holder for scanning the film in the range of 10°–60°.

### FTIR measurements

The CH and NE-CH films were subjected to FTIR by potassium bromide technique in a Nicolet 6700 FT-IR Spectrometer (Thermo Fisher Scientific, Waltham, MA, USA), in the scanning range of 4,000–500  $\text{cm}^{-1}$ . FTIR analysis was done to detect the functional groups present in the sample and to study the interactions between the chitosan and nanoemulsion-impregnated films.

### Antibacterial activity against clinical pathogen *S. aureus*

#### Agar disc diffusion method

The antibacterial activity of CH and NE-CH films (1%, 3%, and 5%) was evaluated by agar disc diffusion method against *S. aureus* Gram-positive bacteria (MTCC 3160; IMTECH, Chandigarh, India). The overnight-grown culture was adjusted to 0.1 optical density (OD) at 600 nm. The plates were previously seeded with 0.1 mL of adjusted culture ( $1 \times 10^7$  CFU/mL). The definite size of the film (5 mm) was kept in the solidified nutrient agar medium in a Petri plate that was incubated for 24 hours at  $37^\circ\text{C} \pm 1^\circ\text{C}$ . Each assay was repeated three times and results were plotted as mean  $\pm$  standard error.

#### Plate count assay – spread plate method

Antibacterial activity of chitosan-only and NE-CH films was evaluated according to a protocol, with a slight modification.<sup>34</sup>

The films (2.5 cm) were transferred to respective test tubes containing 10 mL of 0.1 OD-adjusted *S. aureus* culture. The test tubes were then stored at 28°C–30°C for 24 hours in a shaker. A test tube without film and a test tube without organism were taken as controls. Then, 0.1 mL of each sample was used to prepare serial dilutions that were spread in duplicates on nutrient agar plates. The plates were incubated at 37°C for 24 hours. The colonies were counted and calculated by using a dilution factor. The percentage viability of bacterial cells was plotted after 24 hours of incubation at 37°C. The experiments were done in triplicate and the results were plotted as mean  $\pm$  standard error.

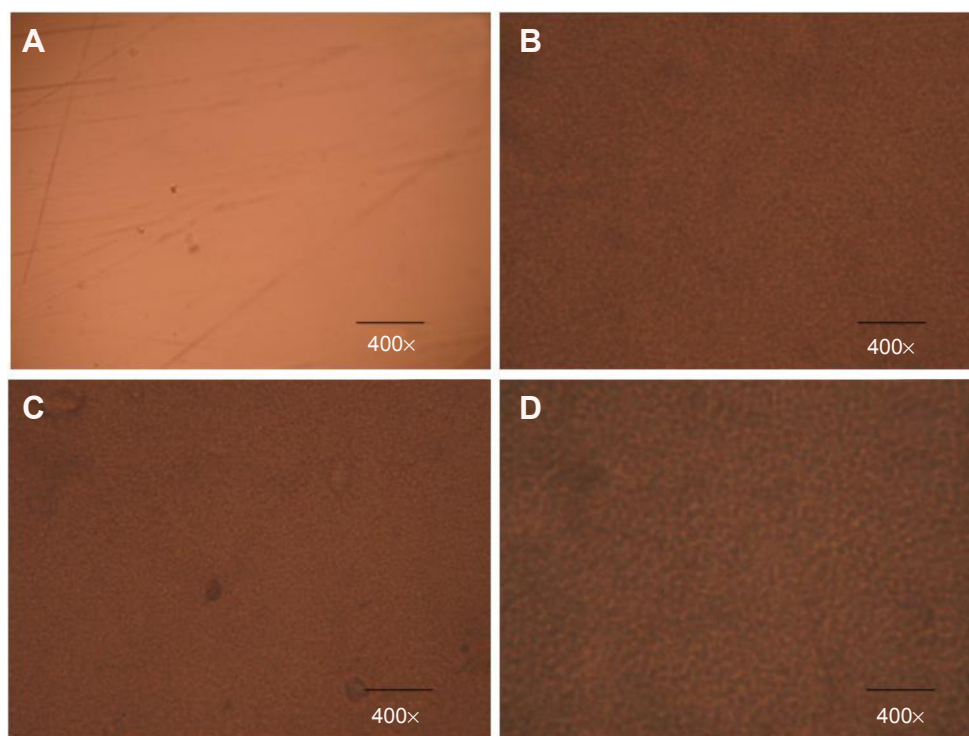
## Results and discussion

### Nanoemulsion formation

The oil-in-water nanoemulsion was formulated using eucalyptus oil and Tween 80 as an organic phase and water as aqueous phase. The volume of 6% oil was used to formulate the 1:2 ratio of the emulsion system. The mean droplet diameter of 9.4 nm with polydispersity index of 0.124 was determined by particle size analyzer. The polydispersity index less than 0.2 indicates the emulsion droplets are monodispersed. The formulated nanoemulsion was stable even after being subjected to centrifugation for 30 minutes at 3,000 rpm. Also, it was more stable when stored at different temperatures (4°C, –20°C, and 45°C) compared to the conventional formulation (before sonication). The impact of non-ionic surfactant type on the droplet size of eucalyptus oil nanoemulsion was studied. In the present work, non-ionic surfactants such as Tween 20 and Tween 80 were used to prepare O/W (oil in water) nanoemulsion due to their high HLB value (16.7 and 15). Eucalyptus oil concentration (6% v/v), oil-surfactant mixing ratio (1:1 v/v), and emulsification time (10 minutes) were kept constant, and nanoemulsions were formulated using different surfactant types (Tween 20 and Tween 80). Of the two surfactants, Tween 80 is more effective in reducing the droplet radii as well as the stability than Tween 20. The low energy emulsification of eucalyptus oil emulsion without sonication was found to be unstable, and phase separated immediately after preparation. Similarly, the results are in agreement with our previous findings, where the nanoemulsion can be formulated with use of non ionic surfactant having droplet radii less than 70 nm using high intensity ultrasonic waves.<sup>37,38</sup>

### Film microstructure

The characteristic optical microscopic images of CH and NE-CH films (1%, 3%, and 5%) are shown in Figure 1. A smooth and continuous microstructure was observed in chitosan film alone (Figure 1A). More oil droplets



**Figure 1** Microstructure of films by phase contrast microscopy at the magnification of 400 $\times$ .

**Notes:** (A) Chitosan-only film. (B) 1% NE-CH. (C) 3% NE-CH. (D) 5% NE-CH.

**Abbreviation:** NE-CH, nanoemulsion-impregnated chitosan film.

were seen with increasing concentration of nanoemulsion and thus were more intimately incorporated into the polymer matrix (Figure 1B–D). A similar morphological structure study was carried out with chitosan film by confocal laser scanning microscopy.<sup>39</sup>

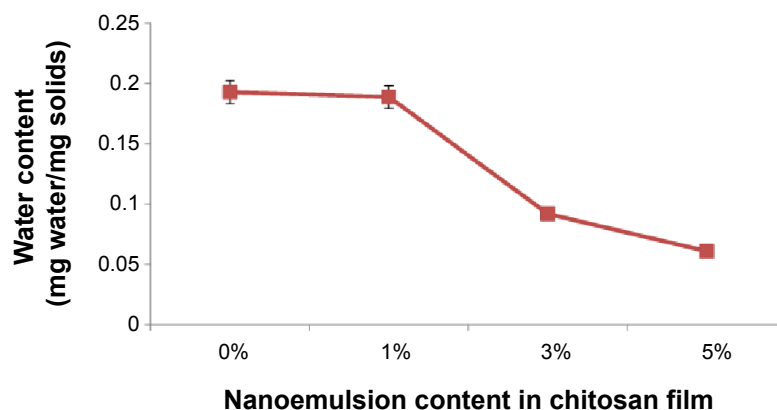
## Moisture content

The hydrophobicity of film provides an indication of resistance to moisture, which is favorable for application studies of pharmaceutical products and in the food and cosmetic industries.<sup>14</sup> Figure 2 shows the variations of the moisture content in

chitosan films for increasing concentrations of nanoemulsion. It can be observed that the moisture content was lowest for films with 5% of emulsion loaded when compared with films having 0%, 1%, and 3% emulsion. This result is similar to that of Shojaee-Aliabadi et al<sup>40</sup> whose films tended to become more hydrophobic due to the oil added to the edible films.

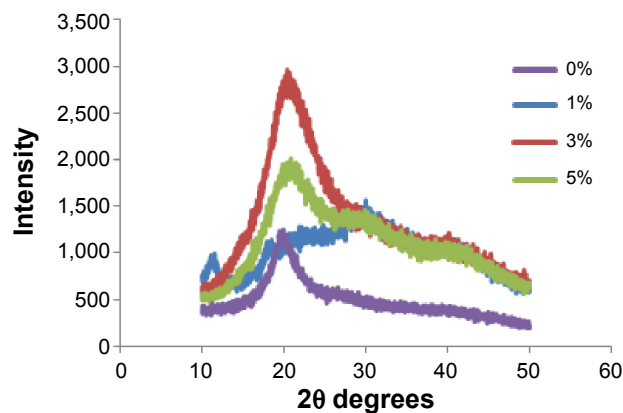
## X-ray diffractogram studies

Chitosan is a polysaccharide that is partially crystalline due to its regular chain and has two reflection falls at  $2\theta=10^\circ$  and



**Figure 2** Moisture content of formulated nanoemulsion-impregnated chitosan films (0%, 1%, 3%, and 5%).

**Note:** Values are given as mean  $\pm$  standard error.

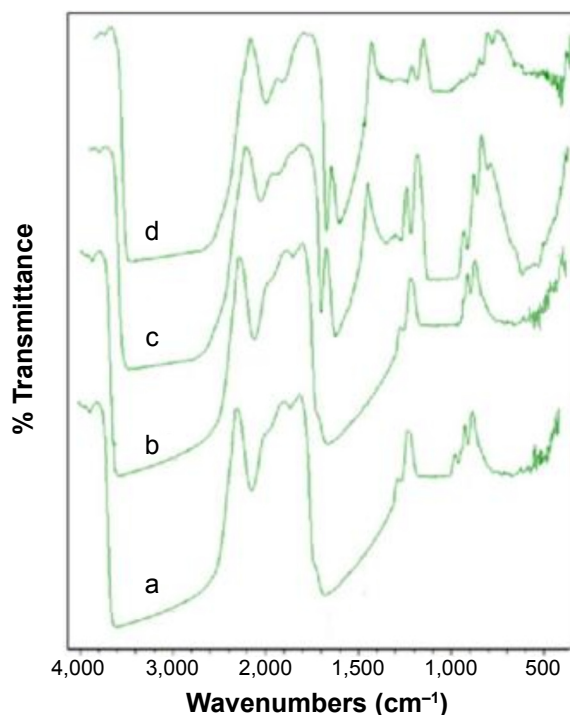


**Figure 3** X-ray diffraction patterns of nanoemulsion-impregnated chitosan films (0%, 1%, 3%, and 5%).

$20^\circ$ .<sup>20</sup> The crystallinity of the chitosan films decreased with the increase of nanoemulsion in films, as shown in Figure 3. The reflection around  $10^\circ$  indicates the presence of the crystal form I. The strongest reflection at  $2\theta=20^\circ$  corresponds to the crystal form II, as per the recent reports by Wu et al<sup>41</sup> and Pastor et al.<sup>42</sup> In emulsion-impregnated films, the peak at  $20^\circ$  became wider, which indicated that the crystallization of chitosan was reduced during the film formation.

### FTIR spectroscopy analysis

FTIR spectroscopy was used to observe the interactions between CH and NE-CH films. Figure 4 shows the



**Figure 4** Fourier transform infrared spectra of NE-CHs.  
**Notes:** (a) 0% NE-CH. (b) 1% NE-CH. (c) 3% NE-CH. (d) 5% NE-CH.  
**Abbreviation:** NE-CH, nanoemulsion-impregnated chitosan film.

FTIR spectra of chitosan and nanoemulsion films (1%, 3%, and 5%). It can be seen that the peak of N-H bending in chitosan and 1% nanoemulsion films located at  $1,668.43\text{ cm}^{-1}$  shifted to  $1,735.93\text{ cm}^{-1}$  with increasing nanoemulsion concentration.<sup>16</sup> The bands appearing between  $2,750$  and  $3,000\text{ cm}^{-1}$  in the spectrum of chitosan film are due to stretching vibrations of C-H bond in  $-\text{CH}_2$  ( $2,930\text{ cm}^{-1}$ ) and  $-\text{CH}_3$  ( $2,870\text{ cm}^{-1}$ ) groups, respectively.<sup>43</sup> The broad band ranging between  $3,500$  and  $3,100\text{ cm}^{-1}$  is attributed to O-H stretching vibration, and the peak at  $1,735\text{ cm}^{-1}$  suggests the presence of a carbonyl group (C=O) in the film. The deconvolution of the FTIR area band ranging between  $1,600$  and  $1,800\text{ cm}^{-1}$  shows that the emulsion incorporation leads to the presence of a new band at  $1,740\text{ cm}^{-1}$ , when nanoemulsion is added. These results are similar when essential oil was added to the chitosan films.<sup>44</sup>

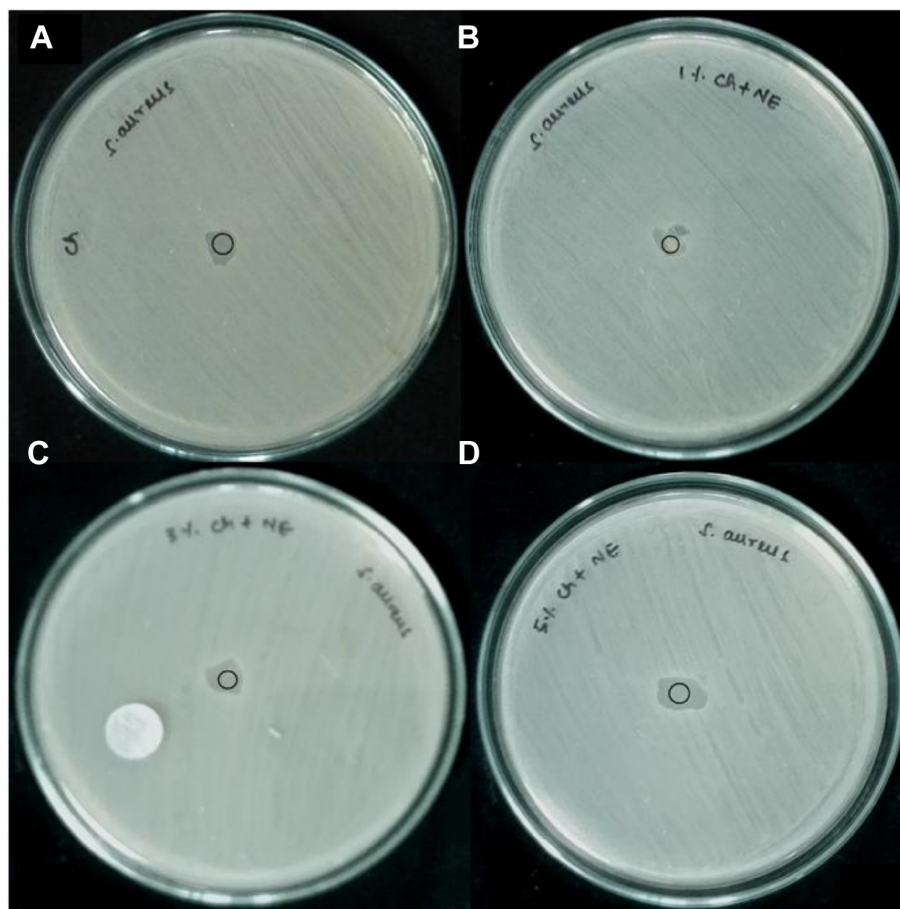
### Antibacterial activity

#### Agar disc diffusion method

The antibacterial activity of CH and NE-CH films (1%, 3%, and 5%) were tested against the tested pathogen *S. aureus* by agar disc diffusion method. The inhibitory activity of chitosan films and NE-CH are shown in Figure 5. The increasing order of 1%, 3%, and 5% NE-CH films (Figure 5B–D) showed higher inhibitory activity (7 mm, 11 mm, and 15 mm, respectively) without any bacterial growth around the film, whereas CH film (Figure 5A) showed 7 mm.

The hydrophilic nature of emulsion-incorporated polymer used in the study was found to have an enhanced inhibitory zone of the tested bacteria (Table 1). The 5% nanoemulsion-impregnated chitosan film showed a higher zone of inhibition than 3%, 1%, and 0% nanoemulsion-impregnated chitosan film. As Hosseini et al mentioned, films containing thyme essential oil in general are very hydrophilic, thus they diffuse faster onto the media with strong inhibition.<sup>45</sup> The increased antibacterial effect of adding essential oil or bioactive compounds on films against Gram-positive and Gram-negative bacteria has been extensively studied.<sup>17,45,46</sup> Positive controls rifampicin, vancomycin, and chloramphenicol were tested against the wound isolate, and the results are shown in Table 2. Rifampicin showed the highest inhibition (29 mm) compared to chloramphenicol (19 mm) and vancomycin (15 mm).

In order to investigate the antibacterial effect of emulsion-impregnated films further, the in vitro quantitative test was examined. The results show that the control films inhibited the growth of the bacteria at a reduced level, which was due to the high molecular weight of the chitosan (90%, deacetylation) that was used. This behavior was also observed by Coma et al



**Figure 5** Antibacterial activity of chitosan-only film and NE-CHs against *Staphylococcus aureus* by agar disc diffusion method on solid medium.

**Notes:** (A) 0% NE-CH. (B) 1% NE-CH. (C) 3% NE-CH. (D) 5% NE-CH.

**Abbreviation:** NE-CH, nanoemulsion-impregnated chitosan film.

wherein high-molecular-weight chitosan did not diffuse through the adjacent agar media.<sup>47</sup>

### Plate count assay

The antibacterial activity of CH film and different concentrations of NE-CH films against the wound isolate *S. aureus* was studied by standard plate count method (Figure 6). Different percentages of NE-CH (1%, 3%, and 5%) with CH films (2.5×2.5 cm) and chitosan-only film (2.5×2.5 cm) were used for the study. As shown in Figure 6E, no viable cells were

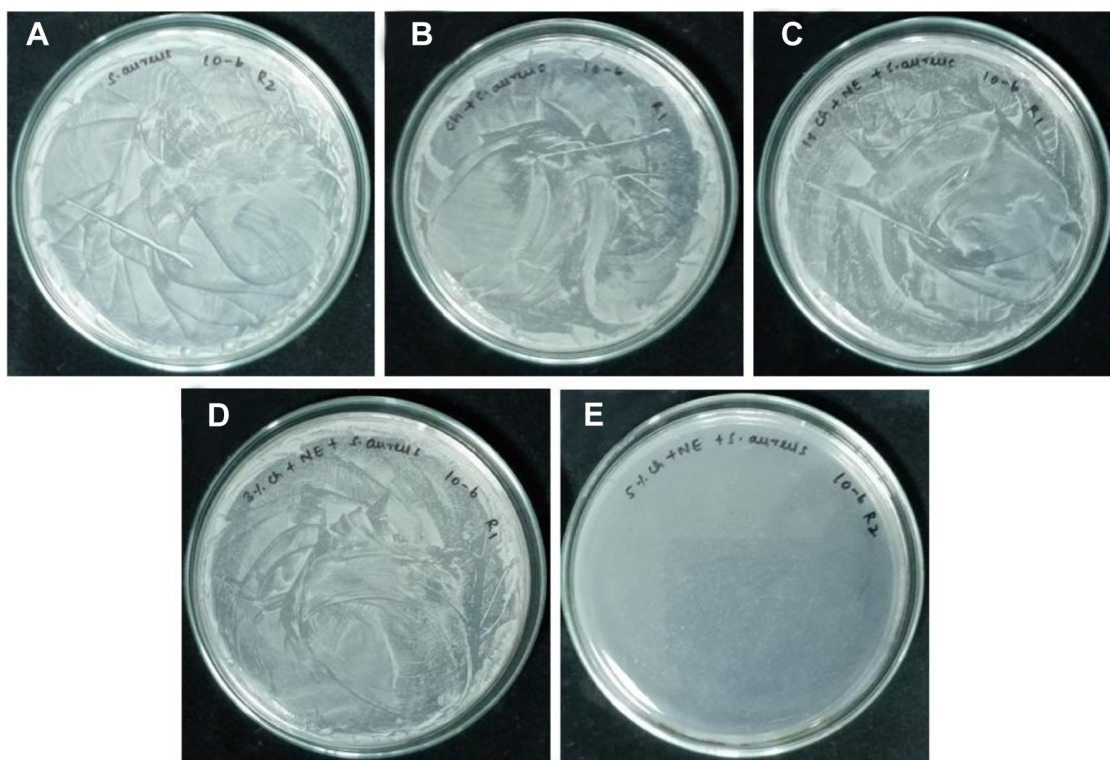
observed after 24-hour treatments with 5% emulsion-impregnated films. For 3% (Figure 6D), 1% (Figure 6C), and chitosan-only film (Figure 6B), 80% viable cells were observed after 24 hours. The control plate (Figure 6A) showed 100% viable cells. It is evident that there is bactericidal activity with the 5% emulsion-impregnated films. The outcome of the study proves that the NE-CH films with increasing concentration of essential oil showed effective antibacterial activity against *S. aureus*. The reduced antibacterial activity was observed for chitosan-only films (Figure 7A) and 1% and 3% NE-CH (Figure 7B and C), due to the biofilm

**Table 1** Antibacterial activity of nanoemulsion-impregnated chitosan films against the wound isolate *Staphylococcus aureus* by agar disc diffusion method

Antibacterial agent	Nanoemulsion content/g chitosan	Inhibitory zone on solid medium (mm)
Nanoemulsion-impregnated chitosan films	0%	7±0.1
	1%	7±0.5
	3%	11±0.3
	5%	15±1

**Table 2** Antibacterial activities of antibiotic discs against the wound isolate *Staphylococcus aureus* by agar disc diffusion method

Positive control antibiotic disc (7 mm)	Zone of inhibition on solid medium (mm)
Vancomycin (30 µg)	15±0.5
Rifampicin (5 µg)	29±0.1
Chloramphenicol (30 µg)	19±0.2



**Figure 6** Antibacterial activity of chitosan-only film and NE-CHs by spread plate method on solid medium.

**Notes:** (A) *Staphylococcus aureus*. (B) 0% NE-CH. (C) 1% NE-CH. (D) 3% NE-CH. (E) 5% NE-CH.

**Abbreviation:** NE-CH, nanoemulsion-impregnated chitosan film.

formation on the inner surface of the test tube (Figure 7), and correlated well with the results of the spread plate method. The turbidity of the bacterial cell was highly reduced when treated with 5% NE-CH film (Figure 7D). The bacteria used in the study were selected as they were found to have

the highest morbidity and mortality associated with wound infections.<sup>48,49</sup>

## Conclusion

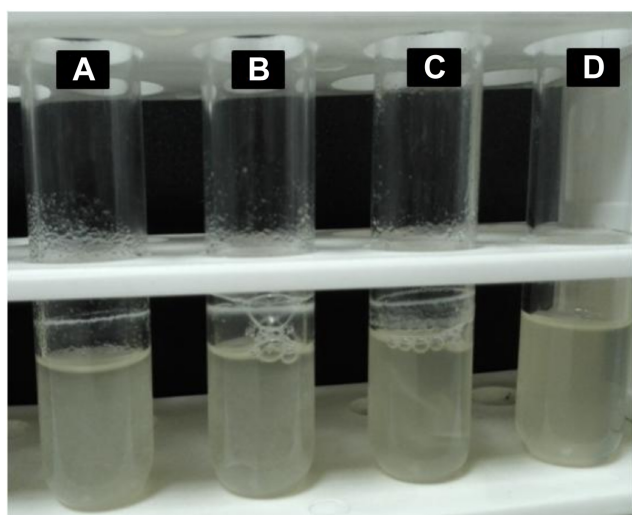
Eucalyptus oil nanoemulsion-impregnated chitosan was successfully developed as a thin film. Further physicochemical parameter measurements were carried out to study the interaction between the chitosan and nanoemulsion films. It was observed that the increasing concentration of nanoemulsion in chitosan film produces reduced moisture content. Also, the NE-CH film was found to have enhanced antibacterial activity against the tested clinical pathogen *S. aureus* than chitosan film alone. These findings could justify the inclusion of NE-CH film for wound management in the pharmaceutical industry.

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## Disclosure

The authors report no conflicts of interest in this work.



**Figure 7** Antibacterial activity of chitosan-only film and NE-CHs by interaction study for 24 hours on liquid medium.

**Notes:** (A) 0% NE-CH. (B) 1% NE-CH. (C) 3% NE-CH. (D) 5% NE-CH.

**Abbreviation:** NE-CH, nanoemulsion-impregnated chitosan film.

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